

**ДЛИТЕЛЬНАЯ АКТИВАЦИЯ ТУЧНЫХ КЛЕТОК КАК
ЭКСПЕРИМЕНТАЛЬНАЯ МОДЕЛЬ ДЛЯ ИССЛЕДОВАНИЯ ИХ РОЛИ
В РЕГУЛЯЦИИ СПЕРМАТОГЕНЕЗА**

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**LONG-TERM ACTIVATION OF MAST CELLS AS AN EXPERIMENTAL
MODEL FOR STUDYING THEIR ROLE IN THE REGULATION OF
SPERMATOGENESIS**

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Резюме

Тучные клетки являются важным компонентом иммунного микроокружения органов мужской репродуктивной системы и участвуют в их регуляции в норме и при патологии. Моделирование дисрегуляции в виде активации или ингибирования тучных клеток и исследование влияния данного нарушения на сперматогенез может помочь в установлении точных механизмов регуляции данного процесса. Одним из активаторов тучных клеток является препарат «Ципрофлоксацин», который зарекомендовал себя в исследованиях тучных клеток сердца, однако ранее не использовался в работах по изучению сперматогенеза.

Цель данного исследования – оценить влияния разных схем приема препарата «Ципрофлоксацин» на тучные клетки репродуктивных органов самцов крыс и выбрать оптимальную дозу и продолжительность приема для создания модели, которая позволит исследовать участие тучных клеток в регуляции сперматогенеза.

Материалы и методы. Эксперимент проведен на самцах линии Wistar. Использовали разные концентрации ципрофлоксацина (200 и 400 мг/кг) и сроки его приема. На гистологических препаратах оценивали морфофункциональные параметры тучных клеток семенников и придатков семенников.

Результаты. Препарат «Ципрофлоксацин» модулирует активность тучных клеток в зависимости от времени, дозы и ткани. После приема препарата в дозе 200 мг/кг 7 суток увеличивается количество, синтетическая активность тучных клеток и процент клеток со зрелыми гранулами как в семенниках, так и в придатках наряду с неизменной дегрануляцией, что указывают на прохождение «подготовительной» фазы, заключающейся в миграции тучных клеток в репродуктивные органы и накоплении ими секрета. После следует начало активной дегрануляции, которая сопровождается возвращением количества тучных клеток к показателю интактной группы, сохранением повышенной синтетической активности и преобладанием в семенниках клеток со зрелыми гранулами. Более высокая доза ципрофлоксацина (400 мг/кг) ускоряет активацию тучных клеток, что приводит к более ранней дегрануляции. Количество и функциональные параметры тучных клеток под действием препарата изменяются аналогично в обоих исследуемых органах, однако наблюдаемые морфометрические изменения и показатели созревания гранул демонстрируют тканеспецифические адаптивные реакции.

Выводы. Проведенное исследование дает основание рекомендовать дозу 400 мг/кг в течение 7 дней для активации тучных клеток в репродуктивных органах самцов крыс и изучения сперматогенеза. Дозу 200 мг/кг следует использовать с целью предварительной стимуляции миграции тучных клеток, повышения их синтетической активности и созревания перед применением другого активатора - индуктора дегрануляции. Данная доза также будет предпочтительной для более длительных экспериментов, чтобы

свести к минимуму потенциальные побочные эффекты, связанные с более высокой дозировкой.

Ключевые слова: тучные клетки, сперматогенез, дегрануляция, активация тучных клеток, ципрофлоксацин, семенник, придаток семенника.

Abstract

Mast cells are an important component of the immune microenvironment in the male reproductive system, involved in both physiological regulation and pathological processes through the secretion of various bioactive substances. Modeling dysregulation via activation or inhibition of mast cells and examining the impact on spermatogenesis can help clarify their exact role in its regulatory mechanisms. Ciprofloxacin, an antibiotic known to activate mast cells, has shown effectiveness for this aim in cardiac mast cell studies but has not been investigated in spermatogenesis. The aim of this study was to evaluate the effects of various ciprofloxacin regimens on mast cells in the reproductive organs of male Wistar rats and determine the optimal dose and duration for creating a model suitable for investigating mast cell involvement in spermatogenesis.

Materials and Methods. Male Wistar rats were treated with ciprofloxacin at 200 and 400 mg/kg for different durations. Morphological and functional characteristics of mast cells in the testes and epididymis were assessed histologically.

Results. Ciprofloxacin modulated mast cell activity in a time, dose, and tissue- dependent manner. A dose of 200 mg/kg for 7 days increased mast cell numbers, enhanced synthetic activity, and raised the proportion of cells with mature granules in both organs, while degranulation remained unchanged. This indicates a "preparatory" phase involving mast cell migration to reproductive tissues and granule accumulation. This was followed by active degranulation after 14 days, associated with return to baseline cell numbers, sustained high synthetic activity, and a predominance of mast cells with mature granules especially in the testes. The higher dose (400 mg/kg) accelerated mast cell activation, leading to earlier degranulation. While functional changes were consistent across both organs, morphometric parameters and granule maturation showed tissue-specific responses. Notably, testicular mast cells displayed minimal morphometric changes, possibly due to the immune - privileged nature of the testes.

Conclusions. Based on these findings, a 400 mg/kg dose for 7 days is recommended to induce mast cell activation for spermatogenesis studies. A 200 mg/kg dose is more suitable for pre-stimulation prior to the use of a degranulation inducer and for long-term studies to minimize potential side effects.

Keywords: mast cells, spermatogenesis, degranulation, mast cell activation, ciprofloxacin, testis, testis appendage.

1 **1 Introduction**

2 Mast cells (MCs) are an important component of the testicular
 3 microenvironment. Under normal conditions, they play a significant role in
 4 maintaining immune privilege and homeostasis within the testicular
 5 microenvironment [4]. This role is largely attributed to the regulatory effects of
 6 MC mediators on vascular permeability and immunomodulation. However, when
 7 MC function is impaired, these effects may shift towards a detrimental influence
 8 on spermatogenesis. Such dysregulation can disrupt the balance of the entire
 9 testicular microenvironment, potentially leading to various pathologies [8].
 10 Modeling MC dysregulation, either through activation or inhibition, and examining
 11 its impact on spermatogenesis can help elucidate the precise mechanisms
 12 governing this process. While immunological factors play a significant role in MC
 13 activation, other various factors, including radiation, pathogens, proteins,
 14 proteolytic enzymes, opioids, estrogens, androgens, and certain antibiotics, can
 15 also trigger their response. Conversely, MC activity can be suppressed by low
 16 doses of γ -ionizing radiation (<0.1 Gy), antihistamines, and MC stabilizers such as
 17 ketotifen [3]. Among antibiotics, fluoroquinolones which are effective against
 18 Gram-positive and Gram-negative bacteria, can also influence MC activity [6].
 19 McNeil and colleagues reported that ciprofloxacin – the most used fluoroquinolone
 20 – can activate MCs via the Mas-related G-protein coupled receptor-X2
 21 (MRGPRX2) [7], a detailed mechanism demonstrated by Liu et al. [5]. While
 22 ciprofloxacin has been used experimentally in studies of cardiac MCs [1], it has not
 23 been previously employed in studies of spermatogenesis, where longer-term
 24 activation is required due to the extended duration of the spermatogenic cycle.

25 The aim of this study is to evaluate the effects of different Ciprofloxacin
 26 administration regimens on MCs in the reproductive organs of male rats and to
 27 determine the optimal dosage and duration for developing a model to study the role
 28 of MCs in the regulation of spermatogenesis.

29 **2 Materials and methods**

30 The experimental animals were sexually mature male Wistar rats, aged 4
 31 months, with a body weight of 350 - 472 g (n=24). During the experiment, the
 32 animals were housed under standard vivarium conditions with a 12-hour light/dark
 33 cycle, without a special diet and with free access to drinking water. The study was
 34 approved by the Ethics Committee of the IIF UB RAS (No. 10-23, dated
 35 09.10.2023).

36 All animals were divided into 4 groups: 1) a group of intact animals (INT),
 37 n=6; 2) a group of animals receiving ciprofloxacin at a dose of 200 mg/kg/day for
 38 7 days (C200D7), n=6; 3) a group of animals receiving ciprofloxacin at a dose of
 39 200 mg/kg/day for 14 days (C200D14), n=6; 4) a group of animals receiving
 40 ciprofloxacin at a dose of 400 mg/kg/day for 7 days (C400D7), n=6.

41 The animals were euthanized by ether overdose, and the testicles and
 42 epididymis were collected. Histological preparations were made and stained with
 43 toluidine blue to assess MC parameters, including their quantity and functional

44 state. Additionally, alcian blue-safranin staining was used to evaluate the degree of
45 MC granule maturity.

46 *Preparation of Drug Suspension.* A ciprofloxacin suspension was prepared
47 by finely crushing two 250 mg ciprofloxacin tablets (Ozon, Russia) and dissolving
48 them in 20 ml of water, resulting in a final concentration of 500 mg/20 ml. The
49 required doses for each animal were calculated and administered via oral gavage.
50 The suspension was thoroughly mixed before each administration to ensure
51 uniform drug distribution.

52 *Calculation of MCs Number.* Scans of prepared tissue sections were obtained
53 using a Leica DM2500 light microscope (Leica, Germany) equipped with a Basler
54 acA1920-40um camera (Basler, Germany) and the MultiMedia Catalog 2008–2020
55 software. The number of MCs was objectively quantified in all scans using QuPath
56 0.5.0 [2], and their density was calculated as the number of MCs per 1 mm² of
57 tissue section.

58 *Assessment of MCs Synthetic Activity.* FIJI ImageJ 1.54f [9] was used to
59 objectively assess MC synthetic activity. The software was calibrated using the
60 Rodbard function to convert grayscale intensity values to optical density (OD).
61 After calibration, all scans were converted to 8-bit grayscale. In each scan, the OD
62 of 50 cells was measured; if fewer than 50 cells were present, all available cells
63 were analyzed. A higher OD value indicates more intense MC synthetic activity.

64 *Assessment of MCs Functional Activity.* MCs were classified into four
65 categories: inactive, weakly degranulating, moderately degranulating, and actively
66 degranulating.

67 The degranulation index (DI) of MCs was calculated using the following
68 formula:

69
$$DI = \frac{D}{D+I} * 100\%$$

70 Where D – the number of MCs with clear signs of degranulation, I – the
71 number of inactive MCs.

72 *Assessment of MCs Granule Maturity.* Using Alcian blue–safranin staining,
73 MCs were classified into three groups:

74 - MCs with immature granules, stained blue due to the affinity of their
75 components for Alcian blue alone (Alc+ granules);

76 - MCs with granules of intermediate maturity, stained purple due to the
77 affinity of their components for both Alcian blue and safranin (Alc+ granules and
78 Saf+ granules);

79 - MCs with mature granules, stained red due to the high content of heparin,
80 a sulfated glycosaminoglycan with a strong affinity for safranin (Saf+ granules).

81 *Evaluation of MCs Morphometric Parameters.* Using FIJI ImageJ 1.54f, the
82 area and perimeter of 50 cells were measured in each preparation. If fewer than 50
83 cells were present, all available cells were analyzed. The software was calibrated
84 by capturing images of an objective micrometer with a known scale. The number
85 of microns per pixel was determined by measuring the distance between marked
86 divisions on the micrometer and inputting this value into the software to ensure
87 accurate morphometric analysis.

88 *Statistical Analysis of the Obtained Data.* All data were analyzed using
 89 GraphPad Prism 9.5.1. The Kruskal-Wallis test was used to compare the number
 90 of MCs, DI, and granule maturity between groups. Dunn's test for pairwise
 91 comparisons, with subsequent correction using the two-stage stepwise method of
 92 Benjamini, Krieger, and Yekutieli, was applied to accurately determine differences
 93 between independent groups.

94 One-way ANOVA was used to compare OD and morphometric parameters
 95 of MCs among three or more groups, followed by the Games-Howell post-hoc test
 96 to determine intergroup differences.

97 Differences were considered statistically significant at $p < 0.05$. Data in the
 98 tables are presented as $M \pm SE$, Me, and the 95% CI of the median.

99 3 Results

100 During the experiment, different dosages and administration durations of the
 101 drug were tested. Administration of ciprofloxacin at 200 mg/kg for 7 days led to an
 102 increase in the number of MCs in the testes and epididymis of rats, with most cells
 103 containing mature granules. An increase in their synthetic activity was observed,
 104 while functional activity remained unchanged (Table 1). These findings suggest
 105 that during the first week of administration, ciprofloxacin stimulates the migration
 106 of MCs to the reproductive organs, enhances their synthesis of bioactive
 107 substances, and promotes granule maturation. However, it does not affect MCs
 108 degranulation, which remains at the same level as intact animals.

109 After 14 days of ciprofloxacin administration at 200 mg/kg, the number of
 110 MCs in the testes and epididymis returns to normal. However, their synthetic
 111 activity remains elevated compared to MCs in intact animals. Unlike the first week,
 112 MCs degranulation in both the testes and epididymis increases significantly
 113 compared to the intact group and the C200D7 group (Table 1).

114 Administration of ciprofloxacin at 400 mg/kg for 7 days produces a
 115 combined effect similar to that of 200 mg/kg administered for 7 and 14 days. The
 116 number of MCs in the testes and epididymis increases significantly, accompanied
 117 by an increase in both synthetic and functional activity compared to the intact group
 118 (Table 1). In the testes, a large number of cells with mature granules are observed,
 119 whereas in the epididymis, there is only a tendency toward an increase in their
 120 number.

121 When comparing MCs parameters in the testes and epididymis, it was found
 122 that the number and functional activity of MCs changed similarly in both
 123 reproductive organs. However, synthetic activity, which generally increased with
 124 drug administration, and the distribution of cells based on granule maturity
 125 exhibited significant organ specificity. In the testes, synthetic activity showed a
 126 dose-dependent increase. In contrast, in the epididymis, synthetic activity
 127 decreased after two weeks of drug administration or when a higher dose of
 128 ciprofloxacin was used. This may suggest that testicular MCs replenish their
 129 granules more rapidly than epididymal MCs following degranulation. Additionally,
 130 in the testes, granule maturation was activated regardless of dose or duration of

131 administration, whereas in the epididymis, this effect was observed only in the
132 C200D7 group.

133 Morphometric characteristics of testicular MCs indicate greater resistance to
134 the effects of ciprofloxacin, as their area remains unchanged compared to the intact
135 group. A small, but significant, increase in their perimeter was observed after the
136 administration of 200 mg/kg ciprofloxacin for two weeks and 400 mg/kg for one
137 week, compared to the C200D7 group but remained at the same level as in the
138 intact group. In contrast, epididymal MCs demonstrate a significant decrease in
139 both area and perimeter following ciprofloxacin administration, regardless of the
140 dose or duration.

141 **4 Discussion**

142 The choice of drug doses in this study is based on the intravenous
143 ciprofloxacin concentration of 150 mg/kg, which has been used in studies of
144 cardiac MCs [1], where stable activation of these cells was demonstrated.
145 According to the Northern Health (Canada) clinical guideline "Intravenous to oral
146 conversion for antimicrobials" the ciprofloxacin conversion factor is 1.25.
147 Therefore, an oral dose of 200 mg/kg was used in this study to activate MCs. To
148 study the effect of an increased dose, a 400 mg/kg dosage (double the initial dose)
149 was administered.

150 The results of this study demonstrate that ciprofloxacin modulates MCs
151 activity in a time-, dose-, and tissue-dependent manner (Fig.1). The increase in the
152 number and synthetic activity of MCs, as well as the activation of their maturation
153 in both the testes and epididymis, observed after administration of the drug at a
154 dose of 200 mg/kg for 7 days, along with unchanged degranulation, suggests the
155 transition through a "preparatory" phase of MCs, characterized by the migration of
156 MCs into the reproductive organs and the accumulation of their secretion. This
157 phase ends in the second week and is marked by the onset of active degranulation
158 and the return of the MCs count to levels seen in the intact group, while synthetic
159 activity remains elevated. Regulatory mechanisms may be involved, initially
160 suppressing MCs degranulation, but eventually being eliminated due to prolonged
161 drug-induced activation.

162 A higher dose of ciprofloxacin (400 mg/kg) accelerates MCs activation,
163 resulting in earlier degranulation. This suggests that MCs response is influenced
164 not only by time but also by dose.

165 While MCs parameters (quantity, synthetic and functional activity) change
166 similarly in the testes and epididymis, the observed morphometric changes -
167 specifically, the reduction in the size and altered shape of epididymal MCs -
168 demonstrate tissue-specific adaptive responses. The absence of significant changes
169 in the morphometric parameters of testicular MCs may be attributed to the unique
170 immune-privileged microenvironment of the testes.

171 The conducted study provides grounds for recommending a dose of 400
172 mg/kg for 7 days to activate MCs in the reproductive organs of male rats and to
173 study spermatogenesis. A dose of 200 mg/kg may be more suitable if the goal is to
174 "prepare" MCs in the reproductive organs by stimulating their migration, synthetic

175 activity, and maturation before applying another activator to induce degranulation.
176 Additionally, this lower dose may be preferable for longer experiments to minimize
177 potential side effects associated with higher dosages.

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ТАБЛИЦЫ

Таблица 1. Влияние ципрофлоксацина на параметры тучных клеток в репродуктивных органах самцов крыс.

Table 1. The effect of Ciprofloxacin on mast cell parameters in the reproductive organs of male rats

Группы Groups	Параметры Parameters							
	Количество ТК на 1 мм ² органа, клетка/мм ² MCs number in 1 mm ² of the organ, cell/mm ²	Синтетическая активность, ОП Synthetic activity, OD	Индекс дегрануляц ии, % Degranulati on index, %	Площадь, мкм ² Area, μm ²	Периметр, мкм Perimeter, μm	ТК с разной степенью зрелости гранул, % Maturation degree of MCs granules, %		
						Незрелые Immature	Промежуто чные Intermediate	Зрелые Mature
Семенник Testis								
ИИТ	0.32 ± 0.04	0.319 ± 0.005	48.647 ± 8.817	38.87 ± 1.56	25.71 ± 0.47	16.99 ± 0.72	23.07 ± 6.56	59.95 ± 6.88
ИИТ	0.30 (0.19; 0.49)	0.330 (0.310; 0.340)	49.45 (15.38; 76.27)	36.00 (32.00; 40.00)	25.35 (24.29; 26.26)	16.67 (15.79; 19.70)	23.07 (0.00; 37.88)	59.95 (42.42; 83.33)

Ц200Д 7	0.94 ± 0.22^a	0.492 ± 0.005^d	32.57 ± 11.03	37.13 ± 0.85	24.45 ± 0.35	2.31 ± 1.33^a	2.26 ± 1.45^a	95.43 ± 1.76^a
C200D 7	0.90 (0.48; 1.77)	0.495 (0.484; 0.503)	21.88 (9.23; 66.67)	37.00 (34.00; 39.00)	23.88 (23.31; 24.97)	1.28 (0.00; 8.33)	0.00 (0.00; 7.69)	95.59 (89.74; 100.00)
Ц200Д 14	0.49 ± 0.07	0.503 ± 0.005^d	73.00 ± 4.88^b	39.30 ± 0.97	26.16 ± 0.38^e	0.91 ± 0.58^a	0.00 ± 0.00^a	99.09 ± 0.58^a
C200D 14	0.51 (0.19; 0.68)	0.511 (0.492; 0.523)	68.51 (61.90; 93.75)	38.00 (35.00; 40.00)	25.89 (24.99; 26.62)	0.00 (0.00; 3.03)	0.00 (0.00; 0.00)	100.00 (96.97; 100.00)
Ц400Д 7	$0.93 \pm 0.18^{a,c}$	$0.533 \pm 0.004^{d,e,f}$	$77.61 \pm 2.31^{a,b}$	37.69 ± 0.82	25.76 ± 0.32^e	0.75 ± 0.34^a	0.24 ± 0.24^a	99.01 ± 0.49^a
C400D 7	0.79 (0.57; 1.80)	0.534 (0.525; 0.544)	80.06 (67.80; 82.00)	36.00 (35.00; 38.00)	25.34 (24.60; 26.32)	0.69 (0.00; 1.69)	0.00 (0.00; 1.43)	99.32 (97.14; 100.00)
Придаток семенника Epididymis								

ИИТ	2.445 ± 0.374	0.389 ± 0.004	54.95 ± 5.59	64.05 ± 2.29	31.67 ± 0.52	33.11 ± 6.53	14.47 ± 2.70	52.42 ± 5.63
ИИТ	2.043 (1.850; 3.817)	0.390 (0.380; 0.410)	55.56 (40.98; 70.97)	56.00 (50.00; 59.00)	30.50 (28.95; 31.65)	30.23 (13.38; 57.89)	12.15 (6.15; 23.94)	46.51 (33.33; 73.85)
Ц200Д 7	9.28 ± 0.89 ^a	0.534 ± 0.004 ^d	37.05 ± 2.19	51.93 ± 1.26 ^d	28.43 ± 0.34 ^d	14.06 ± 3.19	4.86 ± 1.23 ^a	81.08 ± 3.66 ^a
С200Д 7	8.96 (6.89; 12.64)	0.540 (0.530; 0.550)	37.72 (27.47; 43.08)	49.00 (46.00; 51.00)	27.80 (26.97; 28.72)	11.58 (6.14; 27.47)	4.95 (3.00; 1.23)	80.53 (67.03; 91.11)
Ц200Д 14	4.11 ± 0.59 ^b	0.499 ± 0.004 ^{d, e}	71.92 ± 2.61 ^b	55.89 ± 1.41 ^d	29.60 ± 0.36 ^d	22.71 ± 4.62	7.95 ± 1.52	69.33 ± 5.77
С200Д 14	3.82 (2.72; 6.31)	0.510 (0.500; 0.520)	72.08 (62.00; 80.49)	51.50 (48.00; 56.00)	29.23 (28.02; 30.10)	23.49 (8.72; 34.52)	7.98 (2.68; 13.75)	70.89 (52.50; 83.89)
Ц400Д 7	6.21 ± 1.50 ^a	0.492 ± 0.005 ^{d, e}	77.43 ± 3.153 ^{a, b}	55.69 ± 1.48 ^d	29.66 ± 0.38 ^d	18.64 ± 4.15	6.23 ± 1.00 ^a	75.14 ± 4.80
С400Д 7	5.18 (2.63; 11.94)	0.490 (0.480; 0.500)	75.01 (70.18; 91.18)	51.00 (47.00; 55.00)	29.13 (28.16; 29.95)	16.30 (7.32; 35.62)	6.78 (2.44; 8.85)	78.48 (56.85; 90.24)

^a – различия достоверны по сравнению с соответствующим параметром контрольной группы по результатам критерия Краскела–Уоллиса с последующим апостериорным тестом Данна и коррекцией методом Бенджамини, Кригера и Якутиэли ($p < 0,05$)

^b – различия достоверны по сравнению с соответствующим параметром группы Ц200Д7 по результатам критерия Краскела–Уоллиса с последующим апостериорным тестом Данна и коррекцией методом Бенджамини, Кригера и Якутиэли ($p < 0,05$)

^c – различия достоверны по сравнению с соответствующим параметром группы Ц200Д14 по результатам критерия Краскела–Уоллиса с последующим апостериорным тестом Данна и коррекцией методом Бенджамини, Кригера и Якутиэли ($p < 0,05$)

^d – различия достоверны по сравнению с соответствующим параметром контрольной группы по результатам однофакторного дисперсионного анализа (ANOVA) с последующим апостериорным тестом для множественных сравнений по критерию Геймса–Хауэлла ($p < 0,05$)

^e – различия достоверны по сравнению с соответствующим параметром группы Ц200Д7 по результатам однофакторного дисперсионного анализа (ANOVA) с последующим апостериорным тестом для множественных сравнений по критерию Геймса–Хауэлла ($p < 0,05$)

^f – различия достоверны по сравнению с соответствующим параметром группы Ц200Д14 по результатам однофакторного дисперсионного анализа (ANOVA) с последующим апостериорным тестом для множественных сравнений по критерию Геймса–Хауэлла ($p < 0,05$)

^a – differences are significant compared to the corresponding parameter of the control group according to the Kruskal-Wallis test, followed by Dunn post-hoc test, with correction using the Benjamini, Krieger and Yekutieli method ($p < 0.05$)

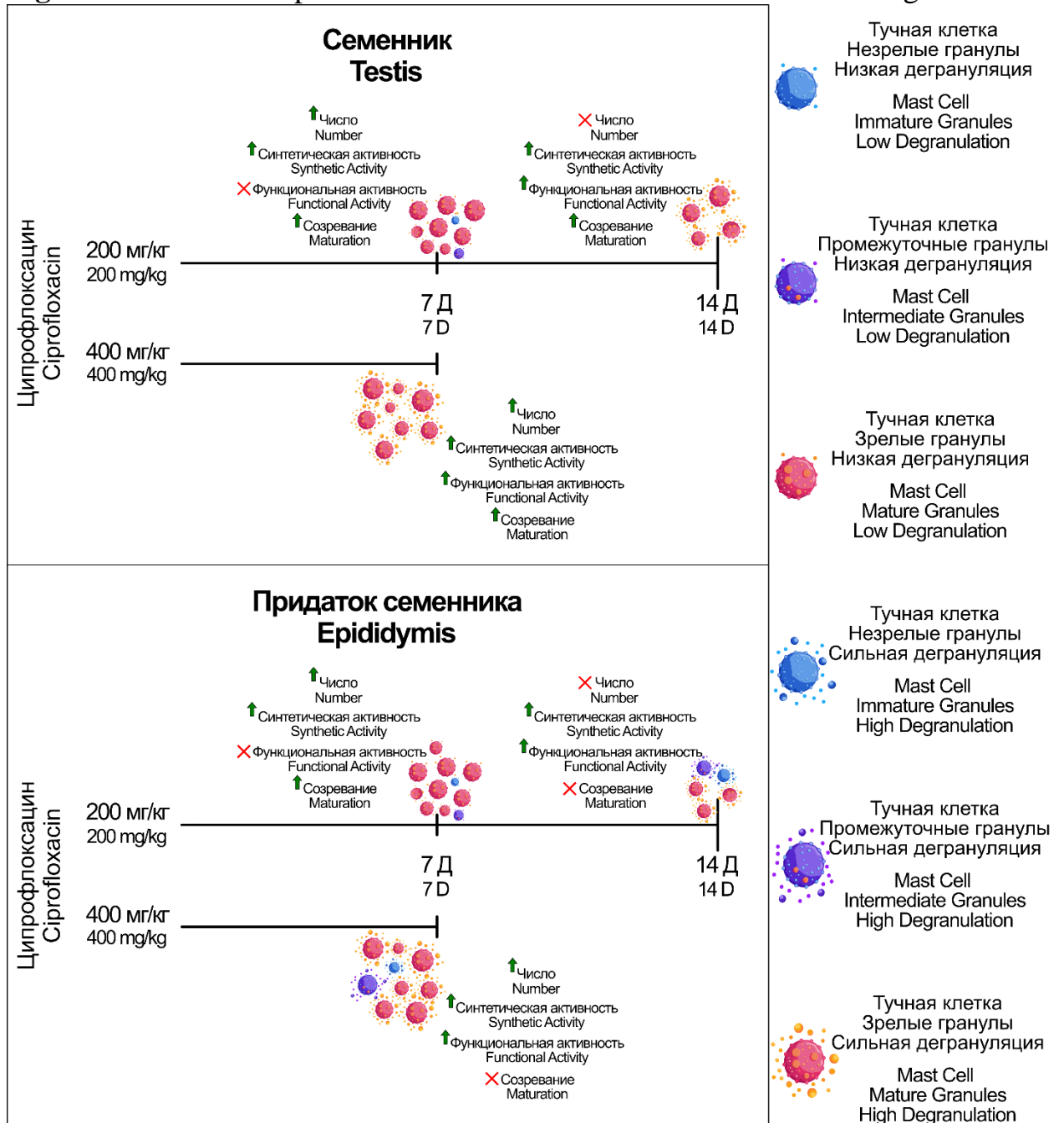
^b – differences are significant compared to the corresponding parameter of C200D7 group according to the Kruskal-Wallis test, followed by Dunn post-hoc test, with correction using the Benjamini, Krieger and Yekutieli method ($p < 0.05$)

- ^c – differences are significant compared to the corresponding parameter of C200D14 group according to the Kruskal-Wallis test, followed by Dunn post-hoc test, with correction using the Benjamini, Krieger and Yekutieli method ($p < 0.05$)
- ^d – differences are significant compared to the corresponding parameter of the control group according to one-way ANOVA with post-hoc test for multiple comparisons using the Games-Howell criterion ($p < 0.05$)
- ^e - differences are significant compared to the corresponding parameter of C200D7 group according to one-way ANOVA with post-hoc test for multiple comparisons using the Games-Howell criterion ($p < 0.05$)
- ^f - differences are significant compared to the corresponding parameter of C200D14 group according to one-way ANOVA with post-hoc test for multiple comparisons using the Games-Howell criterion ($p < 0.05$)

РИСУНКИ

Рисунок 1. Влияние ципрофлоксацина на тучные клетки при разных схемах приема препарата.

Figure 1. Effects of Ciprofloxacin on mast cells with different intake regimens.



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Блок 3. Метаданные статьи

ДЛИТЕЛЬНАЯ АКТИВАЦИЯ ТУЧНЫХ КЛЕТОК КАК
ЭКСПЕРИМЕНТАЛЬНАЯ МОДЕЛЬ ДЛЯ ИССЛЕДОВАНИЯ ИХ РОЛИ В
РЕГУЛЯЦИИ СПЕРМАТОГЕНЕЗА

LONG-TERM ACTIVATION OF MAST CELLS AS AN EXPERIMENTAL
MODEL FOR STUDYING THEIR ROLE IN THE REGULATION OF
SPERMATOGENESIS

Сокращенное название статьи для верхнего колонтитула:

АКТИВАЦИЯ ТУЧНЫХ КЛЕТОК КАК МОДЕЛЬ
MAST CELL ACTIVATION AS A MODEL

Ключевые слова: тучные клетки, сперматогенез, дегрануляция, активация тучных клеток, ципрофлоксацин, семенник, придаток семенника.

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