MOLECULAR AND IMMUNOLOGICAL STUDY OF CELIAC DISEASE IN SAMPLES OF IRAQI PATIENTS

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Abstract

Aim: Evaluation of Human Leukocyte Antigen (HLA) DQ2/DQ8 in celiac disease (CD) patients compared to healthy controls, and to investigate the presence of specific immunological markers (IL-15, IL-21, and TNFα) in the human serum samples from the study groups, also to detect HLA-DQ2 and HLADQ8 in relation to interleukins IL-15, IL-21, and TNF in CD patients.

Methods: A total of 90 individuals participated in this study, comprising 50 patients clinically and serologically diagnosed with celiac disease and 40 apparently healthy individuals serving as the control group. The participants were carefully selected and matched for age and gender where possible. Genotyping for HLA-DQ2 and HLA-DQ8 was conducted using the polymerase chain reaction (PCR) technique, which allows for accurate detection of specific alleles associated with genetic susceptibility to CD. In addition, enzyme-linked immunosorbent assay (ELISA) was employed to determine the serum concentrations of IL-15, IL-21, and TNF-α in both patient and control groups. Statistical analyses were performed to assess the significance of differences observed between the groups and to explore the relationship between HLA genotypes and cytokine expression levels.

Results: The individuals in the case study group were aged between 1 and 60 years. In terms of gender, the patient group consisted of 18 (36.0%) males and 32 (64.0%) females. Among 50 patients with celiac disease, 76.0% had HLA-DQ2, 20.0% had HLA-DQ8, and 14.0% had both. The majority of alleles encoded for HLADQ2 were significant in CD patients when compared with controls. The serum concentrations of IL-15, TNF- α , and IL-21 in the sick group were statistically significant with P-values of 0.001, 0.018, and 0.0001, respectively.

Conclusion: The HLADQ2 genotype is the most common HLA genotype among celiac patients in Iraq, followed by HLADQ8. Serum levels of IL-15, IL-21, and TNF- α were considerably elevated in individuals with CD compared to the control group.

Keywords: Celiac disease, HLA-DQ2, HLA-DQ8, IL-15, IL-21, TNF-α, Iraq, Immunogenetics.

1. Introduction

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Celiac disease, also known as gluten-sensitive enteropathy or non-tropical 2 sprue, is a systemic autoimmune condition that affects genetically predisposed 3 individuals and occurs in about 1% of the global population due to gluten consumption. 4 The immune response is activated by the α -gliadin portion of gluten found in wheat, 5 barley, and rye grains in the small intestine, causing mucosal injury and villous 6 architectural loss. The immune reaction extends beyond the small intestine and can lead 7 to other extraintestinal symptoms throughout the body. Genetic predisposition is crucial 8 in the development of the condition; lifelong gluten intolerance requires the presence of 9 HLA-DQ2 or HLA-DQ8. Having HLA-DQ2 or HLA-DQ8 haplotypes is essential for 10 developing CD. [1] Celiac disease presents with a variety of clinical symptoms. Despite 11 being common, most patients go undiagnosed for an extended period due to the fact that 12 only a small number exhibit the typical malabsorptive signs. [2] Comorbidities include 13 of immune-mediated disorders, dermatitis herpetiformis as a skin symptom, dental 14 enamel hypoplasia (DED), and recurrent oral aphthous lesions (RAS) as oral symptoms. 15 Gluten ataxia is a gluten-induced disorder characterized by the gradual degeneration of 16 Purkinje cells. The common genetic background allows for the simultaneous presence of 17 CD and other autoimmune disorders. Refractory coeliac disease is an uncommon and 18 severe consequence of coeliac disease. Most patients develop ulcerative jejunitis, 19 followed by enteropathy-associated T-cell lymphoma, which virtually exclusively occurs 20 in celiac patients. [3] 21

2. Materials and Methods

The present cross-sectional study aims to analyze the expression levels of specific immunological markers (IL15, IL21, and TNF α) and identify haplotype alterations in HLA typing in patients with CD selected from Medical City Baghdad between November 2022 and February 2023. Fifty patients from the advisory clinic for digestive hospital and the pediatric teaching hospital in the medical city Baghdad participated in the study. The age groupings range from 1 to 60 years. All patients had severe malabsorption, diarrhea and abdominal pain and positively to serological test (anti-tTG Ab and anti-endomysial Ab). Forty controls were included in the study at the premarital examination unit in the medical city Baghdad.

2.1 Human interleukins

Serum human IL-15, IL-21 and TNF α levels measurement, where ELISA kits (CUSABIO\USA) from was used for quantitative determination of human ILs concentration in serum.

2.2 Determination of Celiac disease associated HLA haplotypes using Celiac Strip

HLA genotyping was performed using the polymerase chain reaction-sequence specific oligonucleotide probe (PCR-SSOP) method for HLA-DQ2, HLA-DQ2, HLA-DQ2, HLA-DQ8. The Celiac Strip kit by Operon Immuno & Molecular Diagnostics in Spain can identify the presence or absence of haplotypes encoding HLA-DQ2 and HLA-DQ8, which are the primary HLA haplotypes linked to celiac disease (table 1). The test was carried out in three phases. Genomic DNA was isolated from nucleated cells in a whole blood sample, followed by amplification using PCR techniques. The final step included hybridization and development of the PCR result to identify and evaluate genes. [4]

2.2.1 Extraction and amplification of genomic DNA

The blood samples were processed for genomic DNA extraction following the aseptic conditions outlined in the ReliaPrepTM Blood gDNA Miniprep System methodology by Promega, U.S.A. The DNA samples were amplified using a thermocycler from Analytic Jena GMBH, Germany.

3. Results

Age and Gender Distribution The patient (CD) and control groups were well-matched in terms of age and gender. There was no statistically significant difference in the age distribution (p=0.988), mean age (p=0.560), or gender ratio (p=0.883) between the groups. This means any differences found in later tests are likely due to the disease itself and not because the groups were fundamentally different from the start. (**Table 2**).

Distribution of HLA Haplotypes This table shows the core genetic finding in **Table 3** of the study. A highly significant majority (76%) of celiac patients carried the **HLA-DQ2** gene, compared to only 22.5% of healthy controls (p=0.0001). The **HLA-DQ8** gene was more common in patients (20%) than controls (7.5%), but this difference was not statistically significant (p=0.094). This confirms that HLA-DQ2 is the primary genetic risk factor for celiac disease in this Iraqi population.

HLA Haplotypes by Gender:

Among those who were positive for the risk genes (DQ2, DQ8, or both), there was no significant difference in the distribution between males and females in either the patient or control groups (all p-values > 0.05). This indicates that while the disease is more common in females, the underlying genetic susceptibility is not gender-specific. (Table 4).

Detailed HLA Alleles
This deep dive into the specific components (alleles) of the HLA-DQ2 and DQ8
genes shows that almost all individual alleles (e.g., DQA1*05, DQB1*02,
DRB1*03) were found at a significantly higher frequency in celiac patients than in
controls. This reinforces that the entire HLA-DQ2 haplotype is a strong genetic
marker for the disease (Table 5).
Celiac patients had significantly elevated levels of all three pro-inflammatory
cytokines compared to healthy controls:

- **IL-15:** Much higher in patients (p=0.001).
- **TNF-a:** Significantly higher in patients (p=0.018).
- IL-21: Highly significantly higher in patients (p=0.0001). This indicates a state of intense immune system activation in celiac disease. As shown in Table 6. Among celiac patients, those who carried the HLA-DQ2 gene had significantly
- Among celiac patients, those who carried the **HLA-DQ2** gene had significantly higher levels of **IL-15** (p=0.045) and **TNF-\alpha** (p=0.047) than those who did not carry it. This suggests a link between the main genetic risk factor and a more pronounced

se inflammatory response.(Table 7)

In contrast to DQ2, there was no significant relationship between carrying

- the **HLA-DQ8** gene and the levels of any of the three cytokines in celiac patients.
- This implies that the inflammatory response is more tightly linked to the DQ2
- 91 genotype. (Table 8)
- Table 9 evaluates the potential of these cytokines as diagnostic blood tests for celiac disease:
 - **IL-21** was the best candidate, with high sensitivity (88%), specificity (80%), and overall accuracy (84.4%). This means it is very good at correctly identifying both celiac patients and healthy individuals.
 - **IL-15** showed fair diagnostic performance with 71.1% accuracy.
- TNF- α was a poor standalone diagnostic test with only 58.9% accuracy.

5. Discussion

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5.1 Age and gender distribution of participants.

The results of this study align with findings from several research studies conducted in Iraq and elsewhere. One study on celiac disease in Iraqi patients revealed that 37.5% of the patients were under 10 years old. [5] The age of patients in this study ranged from one year to 60 years, indicating that age-related bias was avoided during patient recruitment. The mean and mode age for all patients and each gender was 19.9±15.4 years. The study results align with previous findings. [6-7] There is no definitive explanation for the age-dependency of celiac disease. However, it is generally observed that the prevalence of celiac disease in children is higher compared to other age groups. This could be attributed to factors such as early introduction of gluten, lack of ongoing breastfeeding, environmental influences during infancy, delayed onset of celiac disease in adulthood, infections, or genetic predisposition. [5,7] Despite using a random sampling technique for patient recruitment, the study population consisted of 64% females and 36% males, resulting in a female to male ratio of 1.9. This occurrence may be explained by the reported female predominance in CD, Hameed and his research colleagues also reported that CD is more in women than in men. [5] Typically, females have a higher prevalence of autoimmune diseases. [8]

5.4 Detection of celiac disease-associated HLA haplotypes (HLA DQ2 and HLA DQ8and HLA DQ2&8) among the patients and the control group.

Multiple studies have shown that the HLA region significantly influences the genetic susceptibility to many systemic autoimmune disorders, such as CD. The HLA class II -DQ area contains genes that encode two alleles, HLA-DQ2 and HLA-DQ8, which are part of the major histocompatibility complex (MHC) II and are associated with hereditary susceptibility to CD .[9] APCs with MHC class II of the HLA-DQ2 or HLA-DQ8 genotype can identify the complex created when gliadin interacts with tissue transglutaminase, leading to immune response activation. This is the foundation of the pathophysiology of CD. [10] Studies from different populations have examined the expression of two genotypes in both individuals with CD and healthy populations. In European region, the prevalence of HLA-DQ2 ranged from 79% to 86% in patients with celiac disease and from 24% to 32% in the general population. For HLA-DQ8, the values were between 10% and 21% in celiac disease patients and 15% to 17% in the general

population. [11-12] In Asia, HLA-DQ2 was found in 80-83% and HLA-DQ8 in around

25% of patients with celiac disease. In the Asian general population, HLA-DQ2 was 133 found in 22-35% of individuals and HLA-DQ8 in 3-22%. [13-15] HLA-DQ2/DQ8 134 heterozygotes have the lowest risk of getting celiac disease, with only a 3% chance 135 despite a prevalence of 25-35% in the general population. The investigation revealed that 136 76.0% of the patients were HLA-DQ2 homozygotes, 20.0% were HLA-DQ8 137 homozygotes, and only 14.0% had HLA-DQ2/DQ8 heterozygosity. In a prior research of 138 74 CD patients, the prevalence of HLA-DQ2 homozygosity was 79.7%, HLA-DQ8 139 homozygosity was 8.1%, and HLA-DQ2/DQ8 was 10.8%. [13] Therefore, the HLA-DQ2 140 homozygosity forms the most predominant genotype in CD patients. [15] The complex 141 molecular mechanism linking HLA-DQ2 homozygosity to the development of CD is not 142 fully understood. However, it is proposed that HLA-DQ2 provides potent antigenic 143 recognition sites for gliadin antigens and has a greater capacity to bind to a wide variety 144 of gluten peptides on the MHC class II of APCs in individuals with this genotype. This 145 may lead to an increased susceptibility to developing gluten-related enteropathy. This 146 study's findings align with the majority of published global investigations, indicating that 147 over ninety percent of individuals with celiac disease exhibit the HLA-DQ2 heterodimer. 148 [17] Also, HLA DQ8 is less strongly associated with CD in the Middle East and North 149 American countries. [18] The results of this study align with those of Çakır and 150 colleagues, who investigated the accuracy of HLA-DQ genotyping and IgA anti-tissue 151 transglutaminase for diagnosing celiac disease in Turkish children. Their findings 152 revealed that 79.3% of celiac children had the DQ2 genotype, while 17.9% had the DQ8 153 genotype. [19] 154 155

5.5 Distribution of HLA haplotypes positive (HLA DQ2, HLA DQ8& HLA DQ2&8) among the study groups according to gender.

To compare the connections between females and males and determine if the parental origin of high-risk haplotypes has an influence. Female patients were shown to have a higher frequency of carrying haplotype DQ2 or DQ8 and DQ2\8 dimers compared to male patients in the study. The correlation between DQ status and gender indicates an influence of the HLA loci on the disease's gender bias. This finding was consistent with the research conducted by Maria C. Mazzilli, B.Sc, in 2007 in Italy and Hameed WS et al. in Iraq. Disagreed with Ahmet Basturk and colleagues in Antalya, Turkey, as well as M. Fernández-Mestre and colleagues in Venezuela on the same topic. [5,17,20]

Detection of CD-associated HLA haplotypes

According to the results of the present study the reports HLA typing information for DQ2 & DQ8 genes in patients with CD and healthy persons firstly regarding the alleles encoded to DQ2, the most common DQA1 alleles were DQA1*05 (82.0%), the most frequent genotype that significantly increased frequency for 41 CD patients, followed by DQB1*02 (78.0%) and DQB1*03 (70.0%) compared to healthy persons (37.5%, 30% and 20%) respectively, results similar to those published. [21-22] This result is similar with Brazilian study by Sliva et al. that revealed statistically significant increase of except DQB1*0302 were the less frequent genotype that the result was no significant P value (0.193). There are studies in regarding this subject that almost agree with the current study. [23] Some investigators have reported in disagreement with such suggestion. [24-25] It is important to consider that several factors such as infections,

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individual dietary habits, genetic make-up, and ethnicity may contribute to this difference.

5.8 Detection of serum IL-15, TNF- α and IL-21 in the patients group and healthy.

IL-15: Serum cytokine elevations in these patients resemble those found in the small intestinal mucosa, while others may originate from sources outside the intestine. Several in vitro investigations have shown that gluten (gliadin) triggers continuous production of pro-inflammatory cytokines of Th type 1. [26] The Th1 response to dietary gluten in the small intestine mucosa is likely responsible for the infiltration of lymphocytes and monocytes in the lamina propria. While having similar effects, Th-1 and Th-2 cytokines mediate distinct functions. The Th-1 response enhances cell-mediated immunity and pro-inflammatory responses, while Th-2 cytokines mainly impact the humoral immune response and help reduce inflammatory processes. Both responses have been noted in CD. [27] The expression of IL-15 varies among different forms of celiac disease, including active, potential, and individuals adopting gluten-free diet. Prior research has verified that individuals with active celiac disease exhibit elevated levels of IL-15 in the mucous membrane of the small intestine. [28] The current investigation found a notably higher amount of IL-15 in patients with CD compared to other groups investigated. Additionally, Abed et al, [27] confirmed the significantly increased level of IL-15 in patients with untreated CD compared with those in the control group. Tamara Vorobjova demonstrated that IL-15 levels were markedly elevated in patients with CD compared to the control group. These levels were also found to be strongly associated with the severity of small intestinal mucosa damage based on the Marsh classification. [27] The current study contradicted Fatemeh Heydari et al's findings in Iran, which demonstrated an insignificant relationship in serum levels between the study groups. [29]

TNF-\alpha: The present investigation found that TNF α levels were higher in patients with effective CD compared to healthy individuals. Gliadin peptides have been demonstrated to stimulate increased TNF-α production by peripheral blood monocytes from Activated CD patients compared to monocytes from Healthy controls or GFD patients in vitro. Similar results have been found worldwide by several estimating methods, including the ELISA method, in countries including Iraq, Iran, and Poland. [27,30] John Sanil Manavalan and colleagues demonstrated that cytokines produced from APC, such as TNF- α, consistently increased in active CD patients compared to those following a gluten-free diet. [26] The USA discovered higher levels of IL15RA and IL-21 expression in duodenal tissues of untreated celiac disease patients compared to controls. IL15 and IL-21 synergistically stimulated intestine intraepithelial cytotoxic T cells. Specifically, they enhanced their transcriptional activity, proliferation, and cytolytic activity. According to a study by Erika Iervasi et al, untreated CD patients have significantly greater IL-21 concentrations in their serum compared to controls, while treated CD patients had low levels of IL-21 [31]. Gluten may act as an autoantigen that stimulates the production of IL-21. Longitudinal research has shown that following a gluten-free diet leads to a significant decrease in the levels of this cytokine. [32-33] IL-21 is known to boost the release of enzymes that break down the extracellular matrix by stromal cells, attractant molecules by epithelial cells, and counteract the immune-

suppressing functions of Treg cells [34]. This could provide additional insight into the

relationship between serum IL-21 levels and mucosal injury. The duodenal mucosa damage progresses gradually over time, culminating in the complete destruction of villi, known as villous atrophy. In this study, for the first time, the possible diagnostic performance of cytokines for CD and the healthy control were evaluated. Our findings demonstrated that IL-21 had the highest sensitivities, specificities, positive and negative predictive values and ACU (84.4) respectively for the detection of the CD patients followed by IL-15 and TNFa and with values ACU (71.1 and 58.9) fair and poor respectively (Table 9).

Correlation of HLA DQ with interleukins (IL-15, TNF-α and IL-21)

There has been a positive correlation between the HLADQ2 of CD patients with IL-15, TNF- α and negative with IL-21(Table 7). Also in the present study, no relation observed between HLADQ8 and immunological markers (cytokines). Histologically, by damage to the intestinal mucosa; and serologically, by the presence of anti-tTG2 Ab, anti-EMA, and/or DGP antibodies. [3] As is well-known, the main genetic determinant in CD involves HLA molecules, specifically the HLADQ2 and/or HLA-DQ8 heterodimers. [35] The physicochemical characteristics of HLA-DQ molecules and their ability to bind certain deamidated peptides by tTG2 are crucial in activating an immune response to gluten, leading to celiac disease. During the adaptive immunological response, gluten-derived peptides that have been altered by tTG2 are displayed by antigen-presenting cells in the mesenteric lymph nodes to CD4 + T cells, in association with HLA-DQ2 and/or DQ8. The Th1-type response results in the generation of IFN- γ and inflammation in the intestines. Earlier researches have shown considerably levels of the serum interleukins were relation with HLA the patients of the CD. [36-37]

At the same time some research has discrepancies these findings. [29,38] this is probably because of the sample size of patients. The conflicting results described by different authors may be due to the heterogeneity of the groups investigated clinical group diversity, the ELISA sensitivity difference, besides differences in the activity indices of the illness, and the variety of management approaches. Probably the main reason due to the small number of positive samples for HLADQ8 in the study.

6. Conclusion

The HLADQ2 genotype is the most common HLA genotype among celiac patients in Iraq, followed by HLADQ8. Celiac disease is more prevalent in children than in other age groups, and it affects females more than males, similar to many autoimmune illnesses. Three proinflammatory cytokines selected for the examination of their role in the immunopathogenesis of CD are IL-15, IL-21, and TNF-α. Each participant exhibited a notable rise in their serum levels among those with CD in comparison to the control group.

AUTHOR CONTRIBUTIONS

All authors have read and approved of the final manuscript.

CONFLICT OF INTEREST

The authors declare that they hold no competing interests.

FUNDING

Self-finding

ТАБЛИЦЫ

Table 1. Haplotypes that encode the HLA-DQ2 and HLA-DQ8

HLA region	Specific haplotype
HLA - DQ2 cis	DQA1*05 - DQB1*02 - DRB1*03
HLA - DQ2 trans	DQA1*05 - DQB1*0301 - DRB1*11/DRB1*12
HP2	DQA1*02 - DQB1*02 - DRB1*07
HLA - DQ8	DQA1*03 - DQB1*0302 - DRB1*04

Table 2. The Age and gender distribution of both patients and control groups.

		Patient CD (N=50)		Control (N=40)		P value		
		No		%	No		%	
	<10years	16		32.0	12		30.0	
	10	14		28.0	10		25.0	
Age (years)	20	8		16.0	7		17.5	0.988
	30	5		10.0	5		12.5	
	=>40years	7		14.0	6		15.0	
	Mean±SD (Range)	19.9±	-15.4	1 (1-60)	21.8±	16.	1 (1-59)	0.560
Gender	Male	18	36.	0	15	37	.5	0.883
	Female	32	64.	0	25	62	5	

^{*}Significant difference between percentages using Pearson Chi-square test (χ^2 -test) at 0.05 level.

Table 3. Distribution of CD-associated HLA haplotypes among the patients and the control group.

HLADQ		Celiac dis	Celiac disease (N=50		N=40)	P value
		No	%	No	%	
	Positive	38	76.0	9	22.5	
DQ2	Negative	12	24.0	31	77.5	0.0001*
	Positive	10	20.0	3	7.5	
DQ8	Negative	40	80.0	37	92.5	0.094
	Positive	7	14.0	2	5.0	
DQ2&DQ8	Negative	43	86.0	38	95.0	0.157

^{*}Significant difference between percentages using Pearson Chi-square test (χ^2 -test) at 0.05 level.

[#]Significant difference between two independent means using Students-t-test at 0.05 level.

Table 4. Distribution of HLA haplotypes positive among the study groups according to gender.

Positive		Celiac	disease	Contro		P value
		No	%	No	%	
	Male	15	39.5	3	33.3	0.733
DQ2	Female	23	60.5	6	66.7	
	Male	3	30.0	2	66.7	0.252
DQ8	Female	7	70.0	1	33.3	
	Male	2	28.6	1	50.0	0.571
DQ2&DQ8	Female	5	71.4	1	50.0	

^{*}Significant difference between percentages using Pearson Chi-square test (χ^2 -test) a 0.05 level.

Table 5. Detection of CD-associated HLA haplotypes among the patients and the control group.

Alleles		Celiac	disease(N=5	Contro	l(N=40)	P value
		No	%	No	%	
DQ2 cis						
	Positive	41	82.0	15	37.5	
DQA1*05	Negative	9	18.0	25	62.5	0.0001*
	Positive	39	78.0	12	30.0	0.0001*
DQB1*02	Negative	11	22.0	28	70.0	
	Positive	35	70.0	8	20.0	0.0001*
DRB1* 3	Negative	15	30.0	32	80.0	
DQ2 trans HP 1						
	Positive	43	86.0	14	35.0	
DQA1*05	Negative	7	14.0	26	65.0	0.0001*
	Positive	23	46.0	10	25.0	
DQB1*0301	Negative	27	54.0	30	75.0	0.040*
	Positive	25	50.0	9	22.5	
DRB1*11	Negative	25	50.0	31	77.5	0.007*
	Positive	9	18.0	-	-	
DRB1*12	Negative	41	82.0	40	100.0	0.005*
DQ2 trans HP2						
DQA1*02	Positive	21	42.0	8	20.0	
	Negative	29	58.0	32	80.0	0.026*
DQB1*02	Positive	31	62.0	10	25.0	
	Negative	19	38.0	30	75.0	0.0001*
DRB1*07	Positive	13	26.0	4	10.0	
	Negative	37	74.0	36	90.0	0.054
DQ8						
DQA1*03	Positive	27	54.0	10	25.0	
	Negative	23	46.0	30	75.0	0.005*

DQB1*0302	Positive	10	20.0	4	10.0	0.193
	Negative	40	80.0	36	90.0	
DRB1*04	Positive	25	50.0	6	15.0	
	Negative	25	50.0	34	85.0	0.001*

^{*}Significant difference between percentages using Pearson Chi-square test (χ^2 -test) at 0.05 level.

Table 6. Detection of serum IL-15, TNF- α and IL-21 in the patients group and healthy.

	Celiac disease (N=50)	Control (N=40)	P value
IL-15 (pg/mL)	15.076±15.076 (1.9	6.666±6.227 (1.12-32.0	0.001#
	78.30)		
TNF-α (pg/mL)	10.406±2.106 (6.7-14.9)	9.451±1.528 (7.42-12.90	0.018#
IL-21 (pg/mL)	22.445±6.547 (14.0-39.0	15.759±4.635 (11.7	0.0001#
	·	31.30)	

#Significant difference between two independent means using Students-t-test at 0.0: level.

Table 7. Relationship between HLA DQ2 with IL-15, TNF- α and IL-21 in patients groups.

	Mean ± SE	Mean ± SE					
DQ2	IL-15 (pg/mL)	TNF- α (pg/mL)	IL-21 (pg/mL)				
Negative	9.153±5.685	9.463±1.736	20.277±6.370				
Positive	16.947±12.626	10.704±2.145	23.130±6.534				
P-value	0.045	0.047	0.191				
#Significant difference between two independent means using Students-t-test at							

#Significant difference between two independent means using Students-t-test at 0.05 level. NS: Non-Significant

Table 8. Relation of HLA DQ8 with interleukins (IL-15, TNF-α and IL-21) among celiac diseases group.

	$Mean \pm SE$					
DQ8	IL-15 (pg/mL)	TNF- α (pg/mL)	IL-21 (pg/mL)			
Negative	15.801±16.227	10.248±2.051	22.103±6.263			
Positive	12.179±9.211	11.040±2.317	23.812±7.799			
T-test	11.41 NS	3.29 NS	4.74 NS			
#Significant difference between two independent means using Students t test at 0.0						

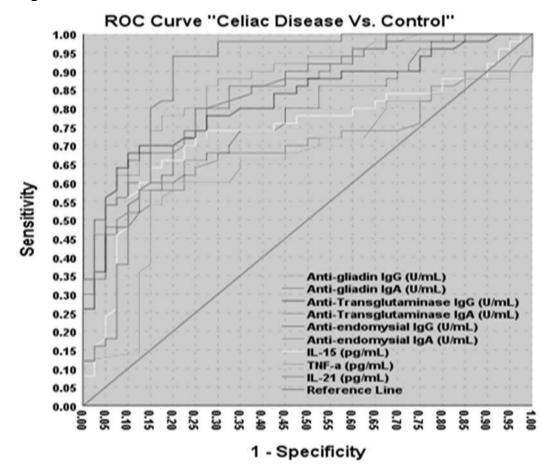
#Significant difference between two independent means using Students-t-test at 0.0: level. NS: Non-Significant.

Table 9. Diagnostic performance of IL-15, TNF- α and IL-21 tests according to the optimal cutoff values for ROC and the manufacturer.

Test Result Variable	Sensitivit	Specificity	PPV	NPV	FN%	FP%	Accuracy rate
IL-15 (pg/mL) \geq 6	74.0	67.5	74.0	67.5	26.0	32.5	71.1
TNF- α (pg/mL) ≥ 9	72.0	42.5	61.0	54.8	25.0	57.5	58.9
IL-21 (pg/mL) ≥17	88.0	80.0	84.6	84.2	12.0	20.0	84.4

РИСУНКИ

Figure 1. ROC curve analysis of IgG and IgA classes for each Ttg, gliadin, endomysial and interleukins (IL-15, TNF- α & IL-21) in celiac disease patients showing area under the curve.



ТИТУЛЬНЫЙ ЛИСТ_МЕТАДАННЫЕ

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Блок 3. Метаданные статьи

MOLECULAR AND IMMUNOLOGICAL STUDY OF CELIAC DISEASE IN SAMPLES OF IRAQI PATIENTS

Сокращенное название статьи для верхнего колонтитула: MOLECULAR AND IMMUNOLOGICAL ASPECTS OF CELIAC DISEASE

Keywords: Celiac disease, HLA-DQ2, HLA-DQ8, IL-15, IL-21, TNF-α, Iraq, Immunogenetics.

Оригинальные статьи. Количество страниц текста – 6, Количество таблиц – 9, Количество рисунков – 1. 12.10.2025

СПИСОК ЛИТЕРАТУРЫ

Reference	Authors, title of a publication	Reference's URL
sequence number		
1.	Voisine J, Abadie V. Interplay Between Gluten, HLA, Innate and Adaptive Immunity	doi:
	Orchestrates the Development of Coeliac Disease. Front Immunol. 2021 Jun	
	2;12:674313.	PMID: 34149709; PMCID: PMC8206552.
2.	Caio G, Volta U, Sapone A, Leffler DA, De Giorgio R, Catassi C, Fasano A. Celiac	doi: 10.1186/s12916-019-138
	disease: a comprehensive current review. BMC Med. 2019 Jul 23;17(1):142.	PMID: 31331324; PMC
		PMC6647104.
3.	Gudjónsdóttir AH, Nilsson S, Naluai AT, Ek J, Amundsen SS, Wahlström J, Ascher H	
	Association between genotypes and phenotypes in coeliac disease. J Pediat	
	Gastroenterol Nutr. 2009 Aug;49(2):165-9.	PMID: 19543113.
4.	Martínez-Ojinaga E, Molina M, Polanco I, Urcelay E, Núñez C. HLA-DQ distribution	
	and risk assessment of celiac disease in a Spanish center. Rev Esp Enferm Dig. 2018	
	Jul;110(7):421-426.	PMID: 29699404.
5.	Smigoc Schweiger D, Mendez A, Kunilo Jamnik S, Bratanic N, Bratina N, Battelino T	
	Brecelj J, Vidan-Jeras B. High-risk genotypes HLA-DR3-DQ2/DR3-DQ2 and DR3	
	DQ2/DR4-DQ8 in co-occurrence of type 1 diabetes and celiac disease. Autoimmunity	-
	2016 Jun;49(4):240-7.	27138053.
6.	Hammo S, Al-Nuaimy WMT, Hayawi M. The significance of CD3 marker in the	- 1 - O
	diagnosis of celiac disease. Ann Coll Med Mosul. 2020;42(2):99–100.	d8/663d28f5161216d535462250
		4439b0239d.pdf

7.	Al-Kemawy SNY, Al-Saltani SNH. Serological tests assessment in patients suspected to	1 01 1
	have celiac disease attending the Gastroenterology and Hepatology Teaching Hospital in	tent/8/Suppl_4/A6.3
	Baghdad. 2024.	1 1 10 100 7 1 10 70 01 1 200
8.	Dixit R, Lebwohl B, Ludvigsson JF, Lewis SK, Rizkalla-Reilly N, Green PH. Celiac	
	disease is diagnosed less frequently in young adult males. Dig Dis Sci. 2014	_
	Jul;59(7):1509-12	24445731
9.	Cecilio LA, Bonatto MW. The prevalence of HLA DQ2 and DQ8 in patients with celiad	
	disease, in family and in general population. Arq Bras Cir Dig. 2015 Jul-Sep;28(3):183	67202015000300009. PM
	5.	26537142; PMCID: PMC47373
10.	Falcigno L, Calvanese L, Conte M, Nanayakkara M, Barone MV, D'Auria G. Structura	doi: 10.3390/ijms21239301. PM
	Perspective of Gliadin Peptides Active in Celiac Disease. Int J Mol Sci. 2020 Dec	33291297; PMCID: PMC77312
	6;21(23):9301.	
11.	Maruntelu I, Preda CM, Sandra I, Istratescu D, Chifulescu AE, Manuc M, Diculescu M	doi: 10.15403/jgld-4187. PM
	Talangescu A, Tugui L, Manuc T, Stroie T, Andrei AC, Tieranu C, Constantinescu I	35694992.
	HLA Genotyping in Romanian Adult Patients with Celiac Disease, their First-degree	
	Relatives and Healthy Persons. J Gastrointestin Liver Dis. 2022 Jun 12;31(2):191-197.	
12.	Fernández-Cavada-Pollo MJ, Alcalá-Peña MI, Vargas-Pérez ML, Vergara-Prieto E	doi: 10.4321/s11
	Vallcorba-Gómez-Del Valle I, Melero-Ruiz J, Márquez-Armenteros AM, Romero	01082013000800005. PM
	Albillos JA, Narváez-Rodríguez I, Fernández-de-Mera JJ, González-Roiz C. Celiad	24274444.
	disease and HLA-DQ genotype: diagnosis of different genetic risk profiles related to the	
	age in Badajoz, southwestern Spain. Rev Esp Enferm Dig. 2013 Sep;105(8):469-76.	
13.	Giriprasad V, Mechenro J, Balamurugan R, Ramakrishna BS. Frequency of HLA celiad	doi: 10.1007/s12664-019-0094
	disease risk alleles and haplotypes in healthy adults in Tamil Nadu. Indian .	Epub 2019 Apr 26. PM
	Gastroenterol. 2019 Apr;38(2):178-182.	31025255.
14.	Özgenel ŞM, Temel T, Üsküdar Teke H, Yıldız P, Korkmaz H, Özakyol A. HLA	doi: 10.5152/tjg.2019.18
	DQ2/DQ8 frequency in adult patients with celiac disease, their first-degree relatives, and	30
	normal population in Turkey. Turk J Gastroenterol. 2019 Apr;30(4):321-325.	PMC6453657.
		1

1.5		1
15.	Mansouri M, Dadfar M, Rostami-Nejad M, Ekhlasi G, Shahbazkhani A, Shahbazkhan	
	B. The frequency of HLA-DQ2/DQ8 haplotypes and celiac disease among the first	
	degree relatives of patients with celiac disease. Gastroenterol Hepatol Bed Bench. 2021	
	Winter;14(1):36-43. PMID: 33868608; PMCID: PMC8035532.	
16.	Talib EQ, Taha GI. Involvement of interlukin-17A (IL-17A) gene polymorphism and	doi: 10.1038/s41405-024-0019
	interlukin-23 (IL-23) level in the development of peri-implantitis. BDJ Open. 2024 Feb	PMID: 38413570; PMC
	28;10(1):12.	PMC10899656.
17.	Basturk A, Artan R, Yilmaz A. The incidence of HLA-DQ2/DQ8 in Turkish children	doi: 10.5114/pg.2017.72099. E
	with celiac disease and a comparison of the geographical distribution of HLA-DQ. Pra	2017 Dec 14. PMID: 293589
	Gastroenterol. 2017;12(4):256-261.	PMCID: PMC5771449.
18.	Siddiqui K, Uqaili AA, Rafiq M, Bhutto MA. Human leukocyte antigen (HLA)-DQ2 and	doi:
	-DQ8 haplotypes in celiac, celiac with type 1 diabetic, and celiac suspected pediatric	
	cases. Medicine (Baltimore). 2021 Mar 19;100(11):e24954.	PMID: 33725967; PMC
		PMC7982179.
19.	Çakır M, Baran M, Uçar F, Akbulut UE, Kaklıkkaya N, Ersöz Ş. Accuracy of HLA-DQ	https://turkjpediatr.org/article/vi
	genotyping in combination with IgA anti-tissue transglutaminase serology and a "scoring	1 91
	system" for the diagnosis of celiac disease in Turkish children. Turk J Pediatr. 2014 Jul	
	Aug;56(4):347-53. PMID: 25818952.	
20.	Fernández-Mestre M, Padrón-Lowe D, Salazar-Alcalá E, Blanco-Pérez F. Role of HLA	doi:
		10.1016/j.rgmxen.2022.03.008.
	Venezuelan population. Rev Gastroenterol Mex (Engl Ed). 2023 Apr-Jun;88(2):125-131	3 6
	venezuelan population. Nev Gustroenteror Wex (Engl Eu). 2023 Tipi Vun,00(2):123 131	35523683.
21.	Martínez-Ojinaga E, Molina M, Polanco I, Urcelay E, Núñez C. HLA-DQ distribution	
21.	and risk assessment of celiac disease in a Spanish center. Rev Esp Enferm Dig. 2018	
	Jul;110(7):421-426	PMID: 29699404
	Jul,110(1).421-420	1 1/1111/1. 47077404

22.	Wiebe C, Kosmoliaptsis V, Pochinco D, Gibson IW, Ho J, Birk PE, Goldberg A	doi: 10.1111/ait 15177 Epub 2
22.	Karpinski M, Shaw J, Rush DN, Nickerson PW. HLA-DR/DQ molecular mismatch: A	
	prognostic biomarker for primary alloimmunity. Am J Transplant. 2019 Jun;19(6):1708	
	1719.	
23.	Maruntelu I, Preda CM, Sandra I, Istratescu D, Chifulescu AE, Manuc M, Diculescu M	doi: 10.15403/jgld-4187. PM
	Talangescu A, Tugui L, Manuc T, Stroie T, Andrei AC, Tieranu C, Constantinescu I	35694992.
	HLA Genotyping in Romanian Adult Patients with Celiac Disease, their First-degree	
	Relatives and Healthy Persons. J Gastrointestin Liver Dis. 2022 Jun 12;31(2):191-197.	
24.	Ramakrishna BS, Venugopal G, Singh A, Pugazhendhi S, Dutta S, Ahuja V, Makharia	doi: 10.1002/jgh3.12651. PM
	GK. Human Leukocyte Antigen DQ (HLA-DQ) genotypes and haplotypes and their	34622007; PMCID: PMC84854
	association with phenotype in patients with celiac disease in India. JGH Open. 2021 Set	
	1;5(10):1190-1196.	
25.	Zamani M, Modares-Sadegi M, Shirvani F, Zamani H, Emami MH. The involvement o	doi: 10.1111/iji.12128. Epub 2
	the HLA-DQB1 alleles in the risk and the severity of Iranian coeliac disease patients. In	Jun 11. PMID: 24917237.
	J Immunogenet. 2014 Aug;41(4):312-7.	
26.	Manavalan JS, Hernandez L, Shah JG, Konikkara J, Naiyer AJ, Lee AR, Ciaccio E	
	Minaya MT, Green PH, Bhagat G. Serum cytokine elevations in celiac disease	· ·
	association with disease presentation. Hum Immunol. 2010 Jan;71(1):50-7.	PMID: 19735687.
27.	Borges MD, Franca EL, Fujimori M, Silva SMC, de Marchi PGF, Deluque AL, Honorio	
	· • • • • • • • • • • • • • • • • • •	10.2174/1871530318666180131
	Cytokines/Chemokines and Adipokines in Serum of Young Adults with Obesity. Endoc	4733. PMID: 29384066.
	Metab Immune Disord Drug Targets. 2018;18(3):260-267.	
28.	Di Sabatino A, Giuffrida P, Fornasa G, Salvatore C, Vanoli A, Naviglio S, De Leo L	
	Pasini A, De Amici M, Alvisi C, Not T, Rescigno M, Corazza GR. Innate and adaptive	-
	immunity in self-reported nonceliac gluten sensitivity versus celiac disease. Dig Live	27130911.
	Dis. 2016 Jul;48(7):745-52.	

29.	Heydari F, Rostami-Nejad M, Moheb-Alian A, Mollahoseini MH, Rostami K	doi:		
2).	Pourhoseingholi MA, Aghamohammadi E, Zali MR. Serum cytokines profile in treated			
	celiac disease compared with non-celiac gluten sensitivity and control: a marker for	30		
	differentiation. J Gastrointestin Liver Dis. 2018 Sep;27(3):241-247.	1 WHD. 30240407.		
30.	Babania O, Mohammadi S, Yaghoubi E, Sohrabi A, Sadat Seyedhosseini F, Abdolahi N	doi: 10.1002/jolo.22094 Epub 2		
50.	Yazdani Y. The expansion of CD14+ CD163+ subpopulation of monocytes and myeloid			
	cells-associated cytokine imbalance; candidate diagnostic biomarkers for celiac disease	FWIC6329136.		
21	(CD). J Clin Lab Anal. 2021 Oct;35(10):e23984.	Ac:		
31.	Iervasi E, Auricchio R, Strangio A, Greco L, Saverino D. Serum IL-21 levels from celiad			
	disease patients correlates with anti-tTG IgA autoantibodies and mucosal damage			
	Autoimmunity. 2020 Jun;53(4):225-230.	Epub 2020 Mar 11. PM		
		32157915.		
32.	Borrelli M, Gianfrani C, Lania G, Aitoro R, Ferrara K, Nanayakkara M, Ponticelli D	30		
	Zanzi D, Discepolo V, Vitale S, Barone MV, Troncone R, Auricchio R, Maglio M. In the			
	Intestinal Mucosa of Children With Potential Celiac Disease IL-21 and IL-17A are Les			
	Expressed than in the Active Disease. Am J Gastroenterol. 2016 Jan;111(1):134-44.			
33.	Galatola M, Izzo V, Cielo D, Morelli M, Gambino G, Zanzi D, Strisciuglio C, Sperandeo	doi: 10.1371/journal.pone.00747		
	MP, Greco L, Auricchio R. Gene expression profile of peripheral blood monocytes:	PMID: 24069342; PMC		
	step towards the molecular diagnosis of celiac disease? PLoS One. 2013 Sep	PMC3775745.		
	17;8(9):e74747.			
34.	Mazzarella G. Effector and suppressor T cells in celiac disease. World J Gastroenterol	doi: 10.3748/wjg.v21.i24.73		
	2015 Jun 28;21(24):7349-56.	PMID: 26139981; PMC		
		PMC4481430.		
35.	Shannahan S, Leffler DA. Diagnosis and Updates in Celiac Disease. Gastrointest Endos	doi: 10.1016/j.giec.2016.08.0		
	Clin N Am. 2017 Jan;27(1):79-92	PMID: 27908520		

36.	Goel G, Daveson AJM, Hooi CE, Tye-Din JA, Wang S, Szymczak E, Williams LJ	doi: 10	.1111/ce	ei.1336	9. Ep	ub 2
	Dzuris JL, Neff KM, Truitt KE, Anderson RP. Serum cytokines elevated during gluten	Oct 1.	PMID:	315050	020; 1	PMC
	mediated cytokine release in coeliac disease. Clin Exp Immunol. 2020 Jan;199(1):68-78	PMC69	004604.			
37.	Ajdani M, Mortazavi N, Besharat S, Mohammadi S, Amiriani T, Sohrabi A, Norouzi A	doi: 10).1186/s	12876-	022-0)245
	Edris G. Serum and salivary tissue transglutaminase IGA (tTG-IGA) level in celia	PMID:	359	33327;	.]	PMC
	patients. BMC Gastroenterol. 2022 Aug 6;22(1):375.	PMC9357310.				
38.	Faghih M, Rostami-Nejad M, Amani D, Sadeghi A, Pourhoseingholi MA, Masotti A, Zal	doi:	10.10)89/gtm	b.201	18.0
	MR. Analysis of IL17A and IL21 Expression in the Small Intestine of Celiac Disease	Epub	2018	Sep	5.	PM
	Patients and Correlation with Circulating Thioredoxin Level. Genet Test Mo	301833	49.			
	Biomarkers. 2018 Sep;22(9):518-525.					